# Plasma Concentrations and Urinary Excretion of Regulatory Peptides in Patients with Urticaria Pigmentosa

Klas Nordlind and Elvar Theodorsson

Department of Dermatology, Academic Hospital, Uppsala, and Department of Clinical Chemistry, Karolinska Hospital, Stockholm, Sweden

#### ABSTRACT

Patients with urticaria pigmentosa were investigated during symptom-free interval regarding plasma concentrations and urinary excretion of immunoreactive regulatory peptides: calcitonin gene-related peptide (CGRP), gastrin, neurokinin A (NKA), neuropeptide Y (NPY), somatostatin (SOM), substance P (SP) and vasoactive intestinal peptide (VIP). The plasma concentrations of these peptides, except for CGRP, were below the detection limit. The urinary excretion of the regulatory peptides were not higher in the patient group than in the controls, but in individual patients there was high urinary excretion of SP and VIP. A lower urinary excretion of CGRP was found in the patient group in addition to a tendency to a lower plasma concentration.

#### INTRODUCTION

In mastocytosis there is a pathologic accumulation of mast cells in various tissues of the body. In most cases the mast cell increase occurs in the skin and the disease is then called urticaria pigmentosa. Common symptoms are pruritus, flushing and whealing, which often develop after strenuous work, sweating, or warm and cold baths, but may also occur spontaneously. Mammalian normal and tumor mast cells have been shown to contain several biologically active substances, including sulfated glycosaminoglycans, histamine, melanin, dopamine, serotonin, arachidonic acid metabolites (for refs see 18, 24) and also VIP

(9). In a previous immunohistochemical investigation (18), somatostatin-like immuno-reactivity was found in skin lesions of patients presenting with urticaria pigmentosa. Some regulatory peptides such as CGRP, opioid peptides, SOM, SP and VIP have been shown to be distributed in neurons in sensory ganglia and/or autonomic ganglia, as well as in peripheral nerve fibers (for refs see e.g. 17). The role of the peptides is not clear, but they may have an inflammatory or trophic effect, as well as functioning as neuromediators by electrophysiological criteria (16).

There is morphologic contact between mast cells and sensory nerves (see e.g. 23), and neuropeptides such as SP (8, 21, 23), VIP and SOM (8) have been shown to release histamine from human cutaneous mast cells.

An increased expression of some regulatory peptides in diseased skin has been reported, e.g. CGRP in prurigo nodularis (1,34), NPY in atopic dermatitis (25), SP in psoriasis (23), atopic dermatitis (22) and prurigo nodularis (1), VIP in eczema (2,21), psoriasis (2,27) and in lichen sclerosus et atrophicus (19), SP and VIP in psoriasis (13) and in bullous and inflammatory skin disease (35), SOM and avian pancreatic polypeptide in diabetic lipodystrophic skin (20), and CGRP, SP and VIP in dermographism and urticaria (36).

As a marker of disease activity in urticaria pigmentosa, the excretion of metabolites of the biogenic amine histamine has been investigated (24). Biogenic amines and regulatory peptides may be colocalized in different tissues. Regulatory peptides are degraded by specific enzymes to biologically inactive metabolites, which may be excreted in the urine.

There have been few reports about urinary excretion of regulatory peptides in normal and pathologic conditions (3,11,30). There might be a possibility of detecting a minor increase in the peptide turnover by analyzing urine samples collected over a 24-h period, since peptide metabolites might be excreted by the kidneys, and antisera as a rule bind to a small part of the peptide, which may be present in the metabolites.

In the present investigation patients with urticaria pigmentosa were

investigated during a symptom-free interval with regard to the plasma concentrations and urinary excretion of various regulatory peptides, namely CGRP, gastrin, NKA, NPY, SOM, SP and VIP, in order to find out whether these patients showed an increased level and excretion of neuropeptides.

## MATERIAL AND METHODS

<u>Patients.</u> Fifteen patients with a mean age of 36 years (range 3-73 years) were admitted to the Department of Dermatology for investigation of urticaria pigmentosa generally affecting the extremities and the trunk. Twenty-six healthy subjects with a mean age of 38 years (5-60 years) were used as control persons.

<u>Plasma</u> samples from six patients and seven control subjects were analyzed.

<u>Urine</u> samples were taken over a 24-h period from 8-15 patients and from 18-26 of the control persons and aliquots of 50 ml were extracted using SepPak (Waters) (31).

<u>Measurement of regulatory neuropeptides.</u> Regulatory peptides were measured by means of competitive radioimmunoassays.

<u>CGRP-like immunoreactivity (CGRP-LI)</u> was assayed using antiserum CGRP8 raised in a rabbit against conjugated rat CGRP. HPLC-purified <sup>125</sup>I-Histidyl rat CGRP was used as radioligand and rat CGRP as standard. The detection limit of the assay for rat CGRP is 9 pmol/L and the cross-reactivity of the assay to SP, NKA, neurokinin B, neuropeptide K, gastrin, neurotensin, bombesin, NPY and calcitonin was less than 0.01%. The cross-reactivities to human CGRP alpha and beta were 93% and 24%, respectively, and to rat CGRP alpha and beta, 100% and 120%, respectively. The intra- and interassay coefficients of variation were 8% and 14%, respectively. <u>Gastrin-LI</u> was assayed with antiserum 4562 (kindly provided by Professor Jens Rehfeld, Rigshospitalet, Copenhagen, Denmark). This antiserum was raised against human non-sulfated sequence 2-17 of gastrin-17. Setting the immunoreactivity for non-sulfated gastrin-17 to 100%, the immuno-reactivity for non-sulfated gastrin-34 is 64%. The antiserum has considerably lower cross-

reactivity to sulfated than to non-sulfated gastrins. Thus the ratio of sulfated to nonsulfated gastrin-34 immunoreactivity is 0.23 (28). The detection limit of the assay was 3.5 pmol/L. The intra- and interassay coefficients were 6% and 11%, respectively. NKA-LI was assayed with antiserum K12, which reacts with NKA (100%), NKA(3-10) (48%), NKA(4-10) (45%), neurokinin B (26%), neuropeptide K (61%) and eledoisin (30%), but not with SP (32). The detection limit of the assay was 12 pmol/L. The intraand interassay coefficients of variation were 7% and 12%, respectively. NPY-LI was assayed with antiserum N1, which cross-reacts 0.1% with avian pancreatic polypeptide, but not with other peptides (33). The detection limit of the assay was 11 pmol/L. The intra- and interassay coefficients of variation were 7% and 12%, respectively. SOM-LI was measured by a competitive immunoassay based on a monoclonal antiserum (Novo Clone SOM-2-28, Novo Bio Labs, Denmark). The detection limit of the assay was 2.3 pmol/L. The intra- and interassay coefficients were 6% and 9%, respectively. SP-LI was assayed with antiserum SP2 (6), which reacts with SP and SP sulfoxide, but not with other tachykinins. The detection limit was 10 pmol/L. The intra- and interassay coefficients of variation were 7% and 11%, respectively. <u>VIP-LI</u> was measured using antiserum 5603-7, kindly provided by Professor Jan Fahrenkrug (15). The detection limit of the assay was 3 pmol/L. The intra- and interassay coefficients of variation were 7% and 11%, respectively.

<u>Statistics.</u> The median and interquartile ranges were used as measures of central tendency and variation, respectively, throughout the study. They will be expressed as follows: median: upper quartile-lower quartile. The Mann-Whitney U test was used for statistical evaluation of results.

## RESULTS

<u>Plasma.</u> The plasma concentrations of gastrin, NKA, NPY, SOM, SP, and VIP were below the detection limits of the assays both in the patient and in the control group. The difference in CGRP between patients and controls was not statistically significant (22+4 and 31+10 (mean+SD) respectively). <u>Urine</u>. In the patient group none of the peptides showed a significantly increased urinary excretion (Fig. 1), but in four patients high excretion values of SP (10, 30, 32 and 74 pmol/day) and in three patients a high excretion of VIP (6, 9 and 13 pmol/day), were observed. In two of these patients, without other clinical symptoms, both these peptides showed elevated values.

A significantly (p<0.01) decreased urinary excretion of CGRP was found in the group of patients (median and quartiles 14.7:8.9-24) compared with the control group (median and quartiles 35:32-37).

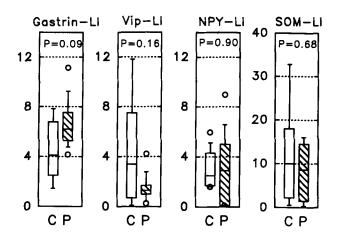
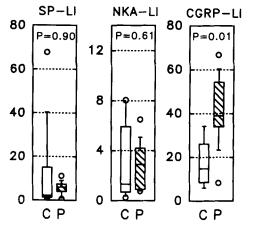


Fig. 1. Box plots of the urinary excretion (pmol/24 h) of immunoreactive regulatory peptidesin patients with urticaria pigmentosa (open boxes) and control subjects (hatched boxes).



### DISCUSSION

An increased production of regulatory peptides has been observed in a wide range of endocrine and nonendocrine tumors. For instance VIP hypersecretion may be a feature of tumors such as pheochromocytoma and hepatocellular carcinoma (7,29), and an elevated concentration of this peptide has been found in the plasma in a case of solitary cutaneous mastocytoma (37).

Most regulatory peptides have a very short half-life in the blood due to rapid degradation (10,14), produced by peptidases. Moreover, mast cells contain proteases that degrade SP and VIP (22). There may also be an indirect mechanism of degradation as an increased level of SP may lead to a decreased level of CGRP as a result of an effect on mast cells and release of proteases degrading CGRP (4,5).

The absence of an increase in urinary excretion of the peptides in this group of patients with urticaria pigmentosa might be due to a short turnover time in the plasma as well as to metabolization of the peptides in the kidneys, and does not rule out the possibility that there may be some increase in the excretion but not sufficiently high to overrule the metabolization rate. As regards CGRP, this peptide might be especially prone to this effect, giving a decreased urinary excretion in the patient group compared with the controls.

Our results might also explain the relative lack of symptoms shown by patients with urticaria pigmentosa, i.e., secreted peptides such as VIP, may undergo rapid degradation.

# ACKNOWLEDGEMENTS

This study was supported by grants from the Swedish Medical Research Council (project 7464) and from the Edvard Welander Foundation. The technical assistance of Kjell Myhr is gratefully acknowledged.

#### REFERENCES

- Abadía Molina, F., Burrows, N.P., Russell Jones, R., Terenghi, G. & Polak, J.M.: Increased sensory neuropeptides in nodular prurigo: a quantitative immunohistochemical analysis. Br J Dermatol 127: 344-351, 1992.
- Anand, P., Springall, D.R., Blank, M.A., Sellu, D., Polak, J.M Bloom, S.R.: Neuropeptides in skin disease: increased VIP in eczema and psoriasis but not axillary hyperhidrosis. Br J Dermatol 124: 547-549, 1991.
- Bonfils, S.: Gastrin-like factor (P.S.U.) in the urine of patients with Zollinger-Ellison syndrome. Physiology of Gastric Secretion, NATO Advanced Study Institute, pp 329-336, 1967.
- Brain, S.D. & Williams, T.J.: Substance P regulates the vasodilator activity of calcitonin gene-related peptide. Nature 335: 73-75, 1988.
- Brain, S.D. & Williams, T.J.: Interactions between the tachykinins and calcitonin gene-related peptide lead to the modulation of oedema formation and blood flow in rat skin. Br J Pharmacol 97: 77-82, 1989.
- Brodin, E., Lindefors, N., Dalsgaard, C.-J., Theodorsson-Norheim, E. & Rosell, S.: Tachykinin multiplicity in rat central nervous system as studied using antisera raised against substance P and neurokinin A. Regul Pept 13: 253-272, 1986.
- Chang, T.M. & Chey, W.Y.: Radioimmunoassay of gastrointestinal peptides in normal and abnormal states. In: Progress in Gastroenterology (ed. G.B.J. Glass & P. Sherlock), vol. 4, pp. 77-132. Grune & Stratton, New York, 1983.
- Church, M.K., Lowman, M.A., Robinson, C., Holgate, S.T. & Benyon, R.C.: Interaction of neuropeptides with human mast cells. Int Arch Allergy Appl Immunol 88: 70-78, 1989.
- Cutz, E., Chan, W., Track, N.S., Goth, A. & Said, S.I.: Release of vasoactive intestinal polypeptide in mast cells by histamine liberators. Nature 275: 661-662, 1978.
- Domschke, S., Domschke, W., Bloom, S.R., Mitznegg, P., Mitchell, S.J. & Lux, G.: Vasoactive intestinal peptide in man-pharmacokinetics, metabolic and circulatory effects. Gut 19: 1049-1053, 1978.

- 11. Du, B., Zhang, J., Eng, J. & Yalow, R.S.: Urinary immunoreactive gastrin in normal subjects. Horm Metab Res 16: 132-135, 1984.
- Ebertz, J.M., Hirshman, C.A., Kettelkamp, N.S., Uno, H. & Hanifin, J.M.: Substance P-induced histamine release in human cutaneous mast cells. J Invest Dermatol 88: 682-685, 1987.
- Eedy, D.J., Johnston, C.F., Shaw, C. & Buchanan, K.D.: Neuropeptides in psoriasis: an immunocytochemical and radioimmunoassay study. J Invest Dermatol 96: 434-438, 1991.
- 14. Fahrenkrug, J.: Evidence for common precursors but different processing of VIP and PHM in VIP producing tumours. Peptides 6: 357-361, 1985.
- Fahrenkrug, J. & Schaffalitzky de Muckadell, O.B.: Radioimmunoassay of vasoactive intestinal polypeptide (VIP) in plasma. J Lab Clin Med 89: 1379-1388, 1977.
- Hartschuh, W., Weihe, E. & Reinecke, M.: Peptidergic (neurotensin, VIP, substance P) nerve fibres in the skin. Immunohistochemical evidence of an involvement of neuropeptides in nociception, pruritus and inflammation.Br J Dermatol 109, suppl 25: 14-17, 1983.
- Johansson, O.: Pain, motility, neuropeptides and the human skin. In: Advances in Pain Research and Therapy Vol. 10. Pain and Mobility (ed. M. Tiengo, A.C. Cuello, J. Eccles & D. Ottoson), pp. 31-44. Raven, New York, 1987.
- Johansson, O. & Nordlind, K.: Immunohistochemical localization of somatostatinlike immunoreactivity in skin lesions from patients with urticaria pigmentosa. Virchows Arch (B) 46: 155-164, 1984.
- Johansson, O. & Nordlind, K.: Immunoreactivity to material like vasoactive intestinal polypeptide in epidermal cells of lichen sclerosus et atrophicus. Am J Dermatopathol 8: 105-108, 1986.
- 20. Johansson, O., Nordlind, K., Efendíc, S. & Lidén, S.: The immuno-histochemical observation of somatostatin- and avian pancreatic polypeptide-like immunoreactivity in certain cellular elements of diabetic lipodystrophic skin. Dermatologica 171: 233-237, 1985.

- 21. Lowman, M.A., Rees, P.H., Benyon, R.C. & Church, M.K.: Human mast cell heterogeneity: histamine release from mast cells dispersed from skin, lung, adenoids, tonsils, and colon in response to IgG-dependent and nonimmunologic stimuli. J Allergy Clin Immunol 81: 590-597, 1988.
- 22. MacQueen, G., Marshall, J., Perdue, M., Siegel, S. & Bienenstock, J.: Pavlovian conditioning of rat mucosal mast cells to secrete rat mast cell protease II. Science 243: 83-85, 1989.
- Naukkarinen, A., Nickoloff, B.J. & Farber, E.M.: Quantification of cutaneous sensory nerves and their substance P content in psoriasis. J Invest Dermatol 92: 126-129, 1989.
- Olafsson, J.H.: Cutaneous and systemic mastocytosis in adults. A clinical, histopathological and immunological evaluation in relation to histamine metabolism. Acta Derm Venereol (Stockh) suppl 115, 1985.
- 25. Pincelli, C., Fantini, F., Massimi, P., Girolomoni, G., Seidenari, S. & Giannetti, A.: Neuropeptides in skin from patients with atopic dermatitis: an immunohistochemical study. Br J Dermatol 122: 745-750, 1990.
- Pincelli, C., Fantini, F., Romualdi, P., Lesa, G. & Giannetti, A.: Skin levels of vasoactive intestinal polypeptide in atopic dermatitis. Arch Dermatol Res 283: 230-232, 1991.
- 27. Pincelli C., Fantini, F., Romualdi, P., Sevignani, C., Lesa, G., Benassi, L. & Giannetti, A.: Substance P is diminished and vasoactive intestinal peptide is augmented in psoriatic lesions and these peptides exert disparate effects on the proliferation of cultured human keratinocytes. J Invest Dermatol 98: 421-427, 1992.
- Rehfeld, J.F., De Magistris, L. & Andersen, B.N.: Sulfation of gastrin: effect of immunoreactivity. Regul Pept 2: 333-342, 1981.
- 29. Said, S.I.: Evidence for secretion of vasoactive intestinal peptide by tumours of pancreas, adrenal medulla, thyroid, and lung: Support for the unifying APUD concept. Clin Endocrinol 5 suppl: 201S-204S, 1976.

- Schwartz, T.W., Saksö, P. & Rehfeld, J.F.: Immunochemical studies on gastrins in the urine. Clin Chim Acta 89: 381-386, 1978.
- 31. Theodorsson-Norheim, E., Hemsén, A. & Lundberg, J.M.: Radioimmuno-assay for neuropeptide Y (NPY): chromatographic characterization of immuno-reactivity in plasma and tissue extracts. Scand J Clin Lab Invest 45: 355-365, 1985.
- Theodorsson-Norheim, E., Norheim, I. & Öberg, K.: Neuropeptide K: a major tachykinin in plasma and tumor tissues from carcinoid patients. Biochem Biophys Res Commun 131: 77-83, 1985.
- 33. Theodorsson-Norheim, E., Hemsén, A., Brodin, E. & Lundberg, J.M.: Treatment of samples when analyzing regulatory peptides. Life Sci 42: 845-848, 1987.
- 34. Vaalasti, A., Suomalainen, H. & Rechardt, L.: Calcitonin gene-related peptide immunoreactivity in prurigo nodularis: a comparative study with neurodermatitis circumscripta. Br J Dermatol 120: 619-623, 1989.
- 35. Wallengren, J., Ekman, R. & Möller, H.: Substance P and vasoactive intestinal peptide in bullous and inflammatory skin disease. Acta Derm Venereol (Stockh) 66: 23-28, 1986.
- 36. Wallengren, J., Möller, H. & Ekman, R.: Occurrence of substance P, vasoactive intestinal peptide, and calcitonin gene-related peptide in dermographism and cold urticaria. Arch Dermatol Res 279: 512-515, 1987.
- 37. Wesley, J.R., Vinik, A.I., O'Dorisio, T.M., Glaser, B. & Fink, A.: A new syndrome of symptomatic cutaneous mastocytoma producing vasoactive intestinal polypeptide. Gastroenterology 82: 963-967, 1982.

Address for reprints: Klas Nordlind Department of Dermatology University Hospital S-751 85 Uppsala